

# **Gene Expression RFP response**

## ***Initial Submission***

EMBL-EBI (European Bioinformatics Institute)

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*OMG Document lifesci/00-03-09 (Gene Expression RFP)*

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## 1. Preface

This submission is in response to LSR RFP, Gene Expression, Object Management Group (OMG) Document lifesci/00-03-09 (Gene Expression RFP)

### 1.1 *Submission Contact Points*

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### 1.2 *Supporting Organisations*

The proposal is supported by the Microarray Gene Expression Database (MGED) group and has been prepared by the Microarray Markup Language (MAML) working group of MGED.

The MGED group is an open discussion group established at the Microarray Gene Expression Database meeting MGED I on November 16-17, 1999, in Cambridge, UK. The goal of the group is to facilitate the adoption of standards for DNA-array experiment annotation and data representation, as well as the introduction of standard experimental controls and data normalization methods. The underlying goal is to facilitate the establishing of gene expression data repositories, comparability of gene expression data from different sources and interoperability of different gene expression databases and data analysis software. Since 1999 the group has had two general meetings and the third one is scheduled for March 28-30, 2001, in Stanford US. MGED group includes representatives from the EMBL-EBI, National Center for Biotechnology Information (NCBI), National Center for Genome Research (NCGR), DNA Databank of Japan (DDBJ), National Human Genome Research Institute, German Cancer Research Centre, Stanford University, University of California at Berkeley, University of Colorado, Rockefeller University, Whitehead Institute, Affymetrix, Incyte and Gene Logic Ltd. MGED has established five working groups, including MAML working group, which is coordinated by Paul Spellman from the University of California at Berkeley (UCLB).

For more information on MGED see <http://www.mged.org/>.

### 1.3 *Acknowledgements*

Below is the list of authors from the MGED MAML working group, who have substantially contributed to the proposal:

Paul Spellman	UCLB	<a href="mailto:spellman@bdgp.lbl.gov">spellman@bdgp.lbl.gov</a>
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The authors would like to thank all the MGED members who have contributed to the proposal.

#### *1.4 Proof of Concept*

MGED group, which includes representatives from most of the major microarray data providers in academia and industry, and major public bioinformatics databases centres, is committed to establishing standards for gene expression profiling.

The EMBL-EBI, NCBI and NCGR are establishing a public repositories for gene expression data which will use the data format proposed in this document. Although currently the data format is based on XML specification, the complete object description will be added in the next submission.

#### *1.5 Response to RFP Requirements*

All the mandatory requirements listed in the items 6.5 of the RFP are fulfilled in this proposal

## **2. Introduction**

We propose a framework for describing information about a DNA-array experiment and a data format – Microarray Markup Language (MAML) – for communicating this information. The information includes details about:

1. Experimental design: the set of the hybridization experiments as a whole;
2. Array design: each array used and each element (spot) on the array;
3. Samples: samples used, the extract preparation and labeling;
4. Hybridizations: procedures and parameters;
5. Measurements: images, quantitation, specifications;
6. Controls: types, values, specifications.

MAML is based on the Extendible Markup Language XML. MAML is independent of the particular experimental platform and provides a framework for describing experiments done on all types of DNA-arrays, including spotted and synthesized arrays, as well as oligo-nucleotide and cDNA arrays, and is independent of the particular image analysis and data normalization methods. MAML does not impose any particular image analysis or data normalization method, but instead provides format to represent microarray data in a flexible way, which allows to represent data obtained from not only any existing microarray platforms, but also many of the possible future variants, including protein arrays. The format allows representation of raw and processed

microarray data. The format is compatible with the definition of the “minimum information about a microarray experiment” (MIAME) proposed by the MGED group, see <http://www.mged.org/>.

The MGED group is an open discussion group initially established at the Microarray Gene Expression Database meeting MGED 1 (November, 1999, Cambridge, UK). The goal of the group is to facilitate the adoption of standards for DNA-array experiment annotation and data representation, as well as the introduction of standard experimental controls and data normalization methods. The underlying goal is to facilitate the establishing of gene expression data repositories, comparability of gene expression data from different sources and interoperability of different gene expression databases and data analysis software.

In the next two sections, we describe the MIAME standard, which describes the content of the information that has to be represented by a data format for microarray gene expression data representation (according to MGED recommendations), followed by the MAML DTD, which defines the actual XML based data format.

### **3. Minimum information about a microarray experiment - (MIAMI)**

Endorsed by MGED steering committee meeting November 17, 2000

The goal of the MIAME is to specify the minimum information that must be reported about a microarray based gene expression monitoring experiment in order to ensure the interpretability of the results and their reproducibility by third parties. The background aim is to help establishing public repositories and data exchange format for microarray based gene expression data. Scientific journals will be encouraged to adopt editorial policies requiring data submissions to repositories, once MIAMI compliant repositories are established.

#### ***Introduction:***

The definition of the minimum information is aimed at cooperative data providers, and not as a legal document meant to close possible loopholes in not providing the information.

Among the concepts in the definition is a list of "qualifier, value, source" triplets, where the "source" is either user defined, or a reference to an externally defined ontology or controlled vocabulary, such as the species taxonomy database at NCBI. Where necessary, the authors are encouraged to define their own qualifiers and provide the appropriate values so that the list as the whole gives sufficient information to interpret the particular part of the experiment. The judgement regarding the necessary level of detail is left to the submitters themselves. In future these 'voluntary' qualifier lists may be gradually substituted by required fields, as the respective ontologies are developed.

Parts of the MIAME can be provided as a reference or link to an externally existing description. For instance, for commercial or other standard arrays all the required information should be normally provided only once by the array provider and referenced by the users. Standard protocols should also normally be provided only once.

#### ***Definition:***

The minimum information about a published microarray based gene expression experiment should include the description of

1. Experimental design: the set of the hybridisation experiments as a whole
2. Array design: each array used and each element (spot) on the array
3. Samples: samples used, the extract preparation and labeling

4. Hybridisations: procedures and parameters
5. Measurements: images, quantitation, specifications
6. Controls: types, values, specifications

The following details should be provided for each array, each sample, hybridisation and measurement in the experiment set:

*1. Experimental design: the set of the hybridisation experiments as a whole*

- a) author (submitter), laboratory, contact information, links (URL)
- b) type of the experiment - maximum one line for instance:
  - normal vs. diseased comparison
  - treated vs. untreated comparison
  - time course
  - dose response
  - effect of gene knock-out
  - effect of gene knock-in (transgenics)
  - shock

(multiple types possible)

- c) experimental factors (e.g., time, dose, genetic variation),
- d) the list of platforms used,
- e) single or multiple hybridisations,

For multiple hybridisations:

- ordered/unordered
  - serial (yes/no)
  - type (e.g., time course, dose response)
  - grouping (yes/no)
  - type (e.g., normal vs. diseased, multiple tissue comparison)
  - list of the samples and arrays used in the experiment and description of the relationship between them: each sample and each array should be assigned a unique id in the experiment set and all the relationships should be listed with appropriate comments
  - which hybridisations are replicates
- f) quality related indicators
    - does a related peer-reviewed publication exist
    - number of replicate hybridisations
    - any other quality control steps taken (polya, unspecific binding etc.)
  - g) optional user defined "qualifier, value, source" list (see Introduction)
  - h) a free text description of the experiment set or a link to a publication

*2. Array design: each array used and each element (spot) on the array.*

- a) array
  - array design name (e.g., "Stanford Human 10K set")
  - platform type: insitu synthesized or spotted

- provider (source)
  - surface type: absorptive/nonabsorptive
  - surface type name
  - array dimensions
  - number of elements on the array
  - a reference system allowing to locate each element (spot) on the array (in the simplest case the number of columns and rows is sufficient)
  - unique ID from the provider
  - production protocol (obligatory if applicable)
  - optional "qualifier, value, source" list (see Introduction)
- b) element (spot) on the array - elements may be simple, i.e., containing only identical molecules, or composite, i.e., containing different oligonucleotides obtained from the same reference molecule; for each element the following must be given:
- position on the array allowing to identify the spot in the image (see 5. a) below);
  - element type: synthesized oligo-nucleotides, PCR products, plasmids, colonies, other;
  - clone information, obligatory for elements obtained from clones:
    - clone ID, clone provider, date, availability
  - sequence information, obligatory for synthetic elements:
    - sequence accession number in DDBJ/EMBL/GenBank if known
    - sequence itself (if databases do not contain it)
    - number of oligos and the reference sequence (or accession number) for multiple oligo-per-element type chips, plus the
    - oligo-sequences, if given
  - approximate lengths if exact sequence not known
  - single or double stranded
  - element (spot) dimensions
  - element generation protocol that includes sufficient information to reproduce the element;
  - gene name and links to appropriate databases (e.g., SWISS-PROT, or organism specific databases), if known and relevant
  - if the element can be used for normalization or control (e.g., element should have expected value)

### 3. *Samples: samples used, extract preparation and labeling*

a) sample source and treatment:

- organism (NCBI taxonomy)
- additional "qualifier, value, source" list; each qualifier in the list is obligatory if applicable; the list includes:
  - cell source and type (if derived from primary sources (s))
  - sex
  - age
  - development stage
  - organism part (tissue)
  - animal/plant strain or line
  - genetic variation (e.g., gene knockout, transgenic variation)
  - individual

- individual genetic characteristics (e.g., disease alleles, polymorphisms)
- disease state or normal
- target cell type
- cell line and source (if applicable)
- in vivo treatments (organism or individual treatments)
- in vitro treatments (cell culture conditions)
- treatment type (e.g., small molecule, heat shock, cold shock, food deprivation)
- compound
- separation technique (e.g., none, trimming, microdissection, FACS)
- laboratory protocol for sample treatment

b) hybridisation extract preparation

- laboratory protocol for extract preparation, including:
  - extraction method
  - whether total RNA, mRNA, or genomic DNA is extracted
  - amplification (RNA polymerases, PCR)
- optional "qualifier, value, source" list (see Introduction)

c) labeling

- laboratory protocol for labelling, including:
  - amount of nucleic acids labeled
  - exogenous sequences (spikes) added
  - label used (e.g., Cy3, Cy5, 33P)
- optional "qualifier, value, source" list (see Introduction)

4. *Hybridisations: procedures and parameters*

- laboratory protocol for hybridisation, including:
  - the solution (e.g., concentration of solutes)
  - blocking agent
  - wash procedure
  - quantity of labelled target used
  - time, concentration, volume, temperature
  - description of the hybridisation instruments
- optional "qualifier, value, source" list (see Introduction)

5. *Measurements: images, quantitation, specifications:*

a) hybridisation scan raw data:

a1) the scanner image file (e.g., TIFF) from the hybridised microarray scanning;

a2) scanning information:

- parsed header of the TIFF file, including laser power, spatial resolution, pixel space, PMT voltage;
- laboratory protocol for scanning, including:
  - scanning hardware



- scanning software
- b) image analysis and quantitation
- b1) the complete image analysis output (of the particular image analysis software) for each element (or composite element - see 2.b)), for each channel;
- b2) image analysis information:
- image analysis software specification and version, availability, and the description of the algorithm
  - all parameters
- c) summarized information from possible replicates
- c1) derived measurement value summarizing related elements as used by the author (this may constitute replicates of the element on the same or different arrays or hybridisations, as well as different elements related to the same entity e.g., gene)
- c2) reliability indicator for the value of c1) as used by the author (e.g., standard deviation); may be "unknown"
- c3) specification how c1 and c2 are calculated; the specification should be based on b1

## *6. Normalisation controls, values, specifications for hybridisations*

- a) Normalization strategy
- spiking
  - "housekeeping gene"
  - total array
  - optional used defined "quality value"
- b) Normalisation algorithm
- linear regression
  - log-linear regression
  - ratio statistics
  - log(ratio) mean/median centering
  - nonlinear regression
  - optional used defined "quality value"
- c) Control array elements
- position (the abstract coordinate on the array)
  - control type (spiking, normalization, negative, positive)
  - control qualifier (endogenous, exogenous)
  - optional used defined "quality value"
- d) Hybridisation extract preparation
- spike type
  - spike qualifier
  - target element
  - optional used defined "quality value"

## 4. MAML DTD

```
<!------->
<!-- MAML DOCUMENT CLUSTER -->

<!ELEMENT maml (analysis_list?,
                array_platform_list?,
                contact_list?,
                creation_info,
                data_set_list?,
                experiment_set_list?,
                hardware_list?,
                protocol_list?,
                sample_list?,
                software_list?,
                publication_list) >

<!------->
<!-- CREATION INFORMATION -->
<!-- A description of the creator of the XML
      document (human, software, hardware) -->

<!ELEMENT creation_info EMPTY >

<!-- date is an ISO date string -->
<!ATTLIST creation_info
            date          CDATA          #REQUIRED
            contact_id    IDREF          #IMPLIED
            software_id    IDREF          #IMPLIED >

<!-- Contact can specify either an individual
      researcher or an organization -->
<!ELEMENT contact_list (contact+) >
<!ELEMENT protocol_list (protocol+) >
<!ELEMENT hardware_list (hardware+) >
<!ELEMENT software_list (software+) >

<!ELEMENT contact (parameter*) >
<!ATTLIST contact
            id            ID            #REQUIRED
            last_name     CDATA         #IMPLIED
            first_name    CDATA         #IMPLIED
            middle_name   CDATA         #IMPLIED
            type          CDATA         #IMPLIED
            lab           CDATA         #IMPLIED
            department    CDATA         #IMPLIED
            organization   CDATA         #IMPLIED
            street        CDATA         #IMPLIED
            city          CDATA         #IMPLIED
            province_state CDATA         #IMPLIED
            country       CDATA         #IMPLIED
            postal_code   CDATA         #IMPLIED
            phone         CDATA         #IMPLIED
            fax           CDATA         #IMPLIED
            email         CDATA         #IMPLIED
            uri           CDATA         #IMPLIED >
```

```

<!-- Can represent PCR, scanner, array printer,
etc. -->
<!ELEMENT hardware (description?,
parameter*) >
<!ATTLIST hardware
id ID #REQUIRED
contact_id IDREF #IMPLIED
type CDATA #IMPLIED
make CDATA #IMPLIED
model CDATA #IMPLIED
serial_number CDATA #IMPLIED
year CDATA #IMPLIED
uri CDATA #IMPLIED >

<!ELEMENT software (description?,
parameter*) >
<!ATTLIST software
id ID #REQUIRED
contact_id IDREF #IMPLIED
hardware_ids IDREFS #IMPLIED
type CDATA #IMPLIED
name CDATA #REQUIRED
version CDATA #IMPLIED
year CDATA #IMPLIED
operating_system CDATA #IMPLIED
uri CDATA #IMPLIED >

<!-- III Protocols -->
<!ELEMENT protocol (standard_protocol,
db_xref?,
protocol_deviations?,
protocol_abstract?) >
<!ATTLIST protocol
id ID #REQUIRED
name CDATA #IMPLIED
type CDATA #IMPLIED >

<!ELEMENT protocol_abstract (#PCDATA) >
<!ATTLIST protocol_abstract
xml:space preserve #FIXED >

<!ELEMENT standard_protocol (#PCDATA) >
<!ATTLIST standard_protocol
xml:space preserve #FIXED >

<!ELEMENT protocol_deviations (#PCDATA) >
<!ATTLIST protocol_deviations
xml:space preserve #FIXED >

<!-- IV Data -->
<!ELEMENT data_set_list (data_set+) >

<!-- for each grouping the first item in the
pair is the rows of the matrix, and the
second element is columns -->
<!ELEMENT data_set ((matrix_axes,
matrix_data),

```

```

                (tagged_data_internal|
                tagged_data_external)) >
<!ATTLIST data_set
    id          ID          #REQUIRED
    name        CDATA      #IMPLIED
    description CDATA      #IMPLIED >

<!ELEMENT matrix_data (ascii_data_internal |
                      ascii_data_external |
                      binary_data_external)* >

<!ELEMENT matrix_axes (matrix_row_list,
                      matrix_column_list,
                      matrix_stack) >
<!ELEMENT matrix_row_list (matrix_row+) >
<!ELEMENT matrix_row EMPTY >
<!ATTLIST matrix_row
    element_id IDREF      #IMPLIED
    image_id   IDREF      #IMPLIED
    quantitation_id IDREF #IMPLIED >

<!ELEMENT matrix_column_list (matrix_column+) >
<!ELEMENT matrix_column EMPTY >
<!ATTLIST matrix_column
    element_id IDREF      #IMPLIED
    image_id   IDREF      #IMPLIED
    quantitation_id IDREF #IMPLIED >

<!ELEMENT matrix_stack (matrix+) >
<!ELEMENT matrix EMPTY >
<!ATTLIST matrix
    element_id IDREF      #IMPLIED
    image_id   IDREF      #IMPLIED
    quantitation_id IDREF #IMPLIED >

<!--
                axis_key refers to either an <element>:id or
                <composite_element>:id; or an <image>:id or
                <composite_image>:id; or a <quantitation>:id or
                <composite_quantitation>:id this is intended to
                reference the missing third dimension of the data
                matrix                                -->

<!--
Data stored internally should be treated as a white
space delimited matrix where null values are specified
as 'NULL'. Carriage returns to delineate the ends of
rows are not necessary.
-->
<!ELEMENT ascii_data_internal (#PCDATA) >
<!ATTLIST ascii_data_internal
    id          ID          #REQUIRED
    type        CDATA      #REQUIRED
    derivation  CDATA      #REQUIRED >

<!--
Data stored externally should be treated as a white
space delimited matrix where null values are specified
as 'NULL'. Carriage returns to delineate the ends of
rows are not necessary.
-->
<!ELEMENT ascii_data_external EMPTY >

```

```

<!ATTLIST  ascii_data_external
            id          ID          #REQUIRED
            type        CDATA       #REQUIRED
            file_uri    CDATA       #REQUIRED >

<!ELEMENT  tagged_data_internal (tagged_data+) >
<!ATTLIST  tagged_data_internal
            id          ID          #REQUIRED
            type        CDATA       #REQUIRED >

<!ELEMENT  tagged_data_external (tagged_data+) >
<!ATTLIST  tagged_data_external
            id          ID          #REQUIRED
            type        CDATA       #REQUIRED >

<!ELEMENT  tagged_data EMPTY >
<!ATTLIST  tagged_data
            element_id  IDREF       #REQUIRED
            image_id    IDREF       #REQUIRED
            quantitation_id IDREF    #REQUIRED
            data        CDATA       #REQUIRED >

<!ELEMENT  binary_data_external EMPTY >
<!ATTLIST  binary_data_external
            id          ID          #REQUIRED
            axis_key    IDREF       #REQUIRED
            type        CDATA       #REQUIRED
            file_uri    CDATA       #REQUIRED >

<!ELEMENT  parameter EMPTY >
<!ATTLIST  parameter
            name        CDATA       #REQUIRED
            value       CDATA       #REQUIRED >

<!-- V      Analysis -->
<!ELEMENT  analysis_list (analysis+) >
<!ELEMENT  analysis (quantitation_list,
                    composite_image_list,
                    composite_quantitation_list,
                    composite_element_list) >

<!--
                    For primary <quantitation> the 'name'
                    should be the same as the column name
                    provided by the scanner software      -->
<!--
                    For a primary <quantitation> the
                    'software_id' really ought to be required
                    and should refer to the scanner software
                    that produced the data                -->
<!ELEMENT  quantitation_list (quantitation+) >
<!ELEMENT  quantitation EMPTY >
<!ATTLIST  quantitation
            id          ID          #REQUIRED
            name        CDATA       #REQUIRED
            software_id IDREF       #IMPLIED
            protocol_id IDREF       #IMPLIED >

<!ELEMENT  composite_element_list (composite_element+) >
<!ELEMENT  composite_element EMPTY >
<!ATTLIST  composite_element

```

```

                id          ID          #REQUIRED
                element_ids IDREFS     #REQUIRED >

<!ELEMENT composite_image_list (composite_image+) >
<!ELEMENT composite_image EMPTY >
<!--
                protocol_id references a protocol that
                describes the method used to create the
                composite elements from the primary
                measurements
-->
<!ATTLIST composite_image
                id          ID          #REQUIRED
                image_ids  IDREFS     #REQUIRED
                protocol_id IDREF      #IMPLIED
                software_id IDREF      #IMPLIED >

<!ELEMENT composite_quantitation_list (composite_quantitation)+>

<!ELEMENT composite_quantitation EMPTY >
<!--
                protocol_id references a protocol that
                describes the method used to create the
                composite from the primary
                measurements
-->
<!--
                We're not sure that software_id is useful
                in this context
-->
<!ATTLIST composite_quantitation
                id          ID          #REQUIRED
                quantitaion_ids IDREFS  #REQUIRED
                protocol_id IDREF      #IMPLIED
                software_id IDREF      #IMPLIED >

<!-- VIII      Experiment Set -->
<!ELEMENT experiment_set_list (experiment_set+) >
<!ELEMENT experiment_set (experimental_design,
                extract_list,
                hybridization_list,
                control_element_list,
                labeled_extract_list,
                sample_list) >

<!ATTLIST experiment_set
                local_accession_number CDATA          #IMPLIED
                experiment_type        CDATA          #IMPLIED
                publication_id          IDREF          #IMPLIED
                contact_id              CDATA          #IMPLIED
                submission_date         CDATA          #IMPLIED
                release_date            CDATA          #IMPLIED
                experiment_date         CDATA          #IMPLIED >

<!ELEMENT experimental_design (biology_description,
                analysis_description,
                experimental_factors,
                quality) >

<!ELEMENT biology_description (#PCDATA) >
<!ATTLIST biology_description
                xml:space preserved          #FIXED >

<!ELEMENT analysis_description (#PCDATA) >
<!ATTLIST analysis_description
                xml:space preserved          #FIXED >

```

```

<!ELEMENT experimental_factors (#PCDATA) >
<!ATTLIST experimental_factors
    xml:space preserved #FIXED >

<!--          We understand that this is limited and
              insufficient, but we believe that quality
              control is an important issue          -->
<!ELEMENT quality (replicates, quality_info*) >
<!ATTLIST quality
    has_replicates (true|false) #REQUIRED >
    peer_reviewed (true|false) false >

<!ELEMENT replicates (description?) >

<!ELEMENT quality_info (#PCDATA) >

<!ELEMENT control_element_list (control_element+) >
<!ELEMENT control_element EMPTY >
<!ATTLIST control_element
    id ID #REQUIRED
    expected_value CDATA #REQUIRED
    quantitation_id IDREF #REQUIRED
    element_id IDREF #REQUIRED >

<!ELEMENT hybridization_list (hybridization+) >
<!ELEMENT hybridization (image+) >
<!ATTLIST hybridization
    name CDATA #REQUIRED
    protocol_ids IDREFS #IMPLIED
    labeled_extract_ids IDREFS #REQUIRED
    control_element_ids IDREFS #REQUIRED
    array_id IDREF #REQUIRED
    id ID #REQUIRED >

<!ELEMENT image EMPTY >
<!ATTLIST image
    protocol_id IDREF #REQUIRED
    labeled_extract_ids IDREFS #REQUIRED
    software_id IDREF #REQUIRED
    hardware_id IDREF #REQUIRED
    file_uri CDATA #REQUIRED
    file_header CDATA #IMPLIED
    microns_per_pixel CDATA #IMPLIED
    image_identifier CDATA #REQUIRED >

<!ELEMENT sample_list (primary_sample|
    derived_sample)+ >

<!ELEMENT derived_sample (treatment+) >
<!ATTLIST derived_sample
    id ID #REQUIRED
    parent_sample_ids IDREFS #REQUIRED >

<!ELEMENT treatment (measurement) >
<!ATTLIST treatment
    action CDATA #REQUIRED
    object CDATA #IMPLIED

```

```

                protocol_id IDREF          #IMPLIED
                order        CDATA         #REQUIRED >

<!-- We don't yet have a full ontology so the primary sample
should include the following kinds of values:

organism_ncbi
organism_other
cell_source
cell_type
sex
age
development_stage
organism_part (tissue)
strain_or_line
genetic_variation
individual
genotype
disease_state
target_cell_type
cell_line_and_source
in_vivo_treatments
in_vitro_treatments
treatment_type
compound
separation_technique          -->
<!ELEMENT  primary_sample (parameter|generic_measure)* >
<!ATTLIST  primary_sample
            id          ID          #REQUIRED >

<!ELEMENT  extract_list (extract+) >
<!ELEMENT  extract (description?,parameter*) >
<!ATTLIST  extract
            id          ID          #REQUIRED
            protocol_id IDREF      #REQUIRED
            type        (total_rna|mrna|dna) #REQUIRED
            sample_ids  IDREFS     #REQUIRED
            label_name  CDATA      #REQUIRED
            name        CDATA      #IMPLIED >

<!ELEMENT  labeled_extract_list (labeled_extract+) >

<!ELEMENT  labeled_extract (description?,parameter*) >
<!ATTLIST  labeled_extract
            id          ID          #REQUIRED
            protocol_id IDREF      #REQUIRED
            extract_ids IDREFS     #REQUIRED
            name        CDATA      #IMPLIED >

<!------->
<!-- DESCRIPTIONS          -->

<!ELEMENT  description  CDATA          >

<!------->
<!-- MEASUREMENT CLUSTER          -->

<!ELEMENT  time          EMPTY >

```



```

<!ATTLIST time
    value CDATA #REQUIRED
    unit (years |
          months |
          weeks |
          d |
          h |
          m |
          s |
          ms |
          us |
          other) #REQUIRED
    other_unit CDATA #IMPLIED >

<!ELEMENT vector (distance+) >
<!ELEMENT distance EMPTY >
<!ATTLIST distance
    value CDATA #REQUIRED
    unit (fm |
          pm |
          nm |
          um |
          mm |
          cm |
          m |
          other) #REQUIRED
    other_unit CDATA #IMPLIED >

<!ELEMENT temperature EMPTY >
<!ATTLIST temperature
    value CDATA #REQUIRED
    unit (C|F) #REQUIRED >

<!ELEMENT mass EMPTY >
<!ATTLIST mass
    value CDATA #REQUIRED
    unit (kg |
          g |
          mg |
          ug |
          ng |
          pg |
          fg |
          other) #REQUIRED
    other_unit CDATA #IMPLIED >

<!ELEMENT volume EMPTY >
<!ATTLIST volume
    value CDATA #REQUIRED
    unit (mL |
          cc |
          dL |
          L |
          uL |
          nL |
          pL |
          fL |
          other) #REQUIRED
    other_unit CDATA #IMPLIED >

```

```

<!ELEMENT concentration EMPTY >
<!ATTLIST concentration
    value CDATA #REQUIRED
    unit (M | mM | uM | nM | pM | fM | mg_per_mL | mL_per_L | g_per_L | g_percent | other) #REQUIRED
    other_unit CDATA #IMPLIED >

<!ELEMENT quantity EMPTY >
<!ATTLIST quantity
    value CDATA #REQUIRED
    unit ( mol | amol | fmol | pmol | nmol | umol | mmol | molecule) #REQUIRED >

<!ELEMENT generic_measure EMPTY >
<!ATTLIST generic_measure
    name CDATA #REQUIRED
    value CDATA #REQUIRED
    unit CDATA #REQUIRED >

<!ELEMENT measurement (time | distance | vector | quantity | temperature | mass | volume | concentration | generic_measure) >
<!ATTLIST measurement
    type (absolute | change) #IMPLIED >

<!------->
<!-- RELATIONSHIPS -->

<!ELEMENT reference (db_xref*,description?) >

<!--
Date is an ISO date string, and is
intended to be used to specify the date
that the reference was made, not the date
the database was released -->

<!ELEMENT db_xref (parameter*) >

```

```

<!ATTLIST db_xref
    database          CDATA          #IMPLIED
    database_version  CDATA          #IMPLIED
    date              CDATA          #IMPLIED
    accession         CDATA          #IMPLIED
    accession_version CDATA          #IMPLIED
    uri              CDATA          #IMPLIED >

<!------->
<!-- PUBLICATION -->

<!ELEMENT publication_list (publication+) >
<!ELEMENT publication (citation | reference) >
<!ATTLIST publication
    id          ID          #REQUIRED >

<!ELEMENT citation (abstract?) >
<!ATTLIST citation
    journal      CDATA          #IMPLIED
    year         CDATA          #IMPLIED
    volume       CDATA          #IMPLIED
    issue        CDATA          #IMPLIED
    page         CDATA          #IMPLIED
    authors      CDATA          #IMPLIED
    publisher    CDATA          #IMPLIED
    editor       CDATA          #IMPLIED
    uri          CDATA          #IMPLIED >

<!ELEMENT abstract (#PCDATA) >

<!------->
<!-- ARRAY PLATFORM -->
<!-- changes: -->
<!-- 1) 'array_def': exchanged 'type' attribute with -->
<!-- 'surface_type' and 'reporter_type' -->
<!-- 2) 'reporter' element converted into Paul's -->
<!-- suggested 'element' element -->

<!ELEMENT array (description?) >
<!ATTLIST array
    id          ID          #REQUIRED
    name        CDATA          #REQUIRED
    array_platform_id IDREF    #REQUIRED >

<!ELEMENT array_platform_list (array_platform|
    array)+ >

<!ELEMENT array_platform (array_def) >
<!ATTLIST array_platform
    id          ID          #REQUIRED >

<!ELEMENT array_def (description?,
    reference*,
    parameter*,
    element*) >

<!ATTLIST array_def
    name        CDATA          #REQUIRED
    contact_id  CDATA          #REQUIRED

```

```

        protocol_id          CDATA          #REQUIRED
        in_situ_synthesis    (true|false)   #REQUIRED
        spotted              (true|false)   #REQUIRED
        surface_type         (non-absorptive|
                             absorptive)    #REQUIRED
        surface_type_name    CDATA          #REQUIRED
        other_surface_type   CDATA          #IMPLIED
        number_of_elements   CDATA          #IMPLIED
        short_axis_length    CDATA          #IMPLIED
        long_axis_length     CDATA          #IMPLIED
        element_type         (single-multimer |
                             multiple-oligomer |
                             other)         #REQUIRED
        model_name           CDATA          #IMPLIED
        version              CDATA          #IMPLIED
        uri                  CDATA          #IMPLIED >

<!------->
<!-- ELEMENT ----->
<!ELEMENT element          ((bio_seq|
                             ref_bio_seq|
                             ref_clone)+,
                             gene*,
                             parameter*,
                             measurement*,
                             description?) >

<!--          sequence_length can be approximate          -->
<!--          diameter can be approximate                 -->
<!--          empty elements should have an empty        -->
<!--          <bio_seq>                                   -->

<!ATTLIST element
        id          ID          #REQUIRED
        attachment_method CDATA  #IMPLIED
        strandedness (single|double) #IMPLIED
        type        (empty|
                    pcr|
                    synthesized_oligo|
                    intact_plasmid|
                    colony)      #REQUIRED
        diameter    CDATA        #IMPLIED
        sequence_length CDATA    #IMPLIED
        location     CDATA        #IMPLIED
        protocol_id  CDATA        #IMPLIED
        row          CDATA        #IMPLIED
        column       CDATA        #IMPLIED
        block        CDATA        #IMPLIED
        x_microns    CDATA        #IMPLIED
        y_microns    CDATA        #IMPLIED
        name         CDATA        #IMPLIED >

<!ELEMENT bio_seq          (#PCDATA|db_xref) >
<!ELEMENT ref_bio_seq     (#PCDATA|db_xref) >
<!ELEMENT gene             (#PCDATA|db_xref) >
<!ELEMENT ref_clone       (#PCDATA|db_xref) >

```